Growth inhibitory efficacy and anti-aromatase activity of *Tabebuia avellanedae* in a model for post-menopausal Luminal A breast cancer

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Abstract. Aromatase inhibitors (AIs) represent a treatment option for post-menopausal estrogen receptor-positive (ER⁺) breast cancer as monotherapy, or in combination with cyclin-dependent kinase 4/6 or mTOR inhibitors. Long-term treatment with these agents leads to dose-limiting toxicity and drug resistance. Natural substances provide testable alternatives to current therapy. *Tabebuia avellanedae* (TA) tree is indigenous to the Amazon rainforest. The inner bark of TA represents a medicinal dietary supplement known as Taheebo. Non-fractionated aqueous extract from TA is an effective growth inhibitor in the Luminal A and triple negative breast cancer models. The quinone derivative naphthofurandione (NFD) is a major bioactive agent in TA. The present study examined the efficacy of finely ground powder from the inner bark of TA, available under the name of Taheebo-NFD-Marugoto (TNM). The ER⁺ MCF-7 cells stably transfected with the aromatase gene *MCF-7AROM* represented a model for aromatase-expressing post-menopausal breast cancer. Anchorage-independent colony formation, cell cycle progression, pro-apoptotic caspase 3/7 activity, apoptosis-specific gene expression, aromatase activity and select estradiol (E₂) target gene expression represented the mechanistic end points. Treatment of MCF-7AROM cells with TNM induced a dose-dependent reduction in E₂-promoted anchorage-independent colony number. Mechanistic assays on TNM-treated MCF-7AROM cells demonstrated that TNM at a concentration of 10 µg (NFD content: 2 ng), induced S-phase arrest, increased pro-apoptotic caspase 3/7 activity, increased pro-apoptotic BAX and decreased anti-apoptotic *BCL-2* gene expression, and inhibited aromatase activity. Additionally, TNM treatment downregulated *ESR-1* (gene for ER-α), aromatase and progesterone gene expression and reduced mRNA levels of E₂ target genes *pS2*, *GRB2* and *cyclin D1*. Inhibition of aromatase activity, based on the NFD content of TNM was superior to the clinical AIs Letrozole and Exemestane. These data demonstrated the potential efficacy of TNM as a nutritional alternative for current therapy of aromatase positive, post-menopausal breast cancer.

Introduction

Clinical treatment options for post-menopausal hormone receptor positive, human epidermal growth factor receptor-2 negative breast cancer include the use of selective estrogen receptor modulators and aromatase inhibitors (1, 2). Aromatase inhibitor based monotherapy is frequently combined with selective small molecule inhibitors of CDK4/6 and mTOR pathways. Long-term treatment involving single agent therapy or multi-agent combination therapy is frequently associated with acquired drug resistance predominantly due to the emergence of cancer stem cells, thereby impacting therapeutic efficacy and promoting disease progression (3-5).

Aromatase CYP19 A1 functions as a critical enzyme for peripheral and intra-tumoral estrogen bio-synthesis via conversion of adrenal androstenedione to estrone (E₁) and subsequently to estradiol (E₂) thereby providing growth-promoting estrogens. Pharmacological agents Letrozole (LET) and Exemestane (EXM) are selective inhibitors of aromatase (1, 2). These agents exhibit acquired tumor resistance in preclinical models for aromatase-expressing Luminal A breast cancer, as well as in estrogen receptor-positive clinical breast cancer (6-12).

Naturally occurring non-toxic substances including dietary supplements and natural botanicals are widely used in complemental and alternative medicine. Natural products exhibiting effective inhibition of aromatase activity may represent potential testable alternatives to the limitations of clinical aromatase inhibitors. *Tabebuia avellanedae* (TA) is a tree native to the Amazon rainforest. Drinks from the bark of the TA tree have been traditionally used by the indigenous population to address a wide variety of health issues. A non-fractionated powder from

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the inner bark of TA, under the name Taheebo, is available from Taheebo Japan, Co., Ltd.. An aqueous extract of Taheebo has been documented to exhibit anti-cancer activity in animal models for organ site cancers (13), as well as growth inhibitory efficacy in human carcinoma-derived cell culture models for prostate and breast cancer via multiple mechanisms (14-16). Growth inhibitory efficacy of TA extract in a model for Luminal A breast cancer subtypes is associated with the differential expression of proliferation and apoptosis-specific genes (17). In a model for triple-negative breast cancer the inhibitory efficacy of TA is attributable to inhibition of G0 to S phase transition, induction of pro-apoptotic caspase 3/7 activity and modulation of the RB pathway (18).

Recently, Taheebo Japan invented a proprietary process capable of producing TA powder that is considerably more finely ground than the original TA powder. The new product marketed by Taheebo Japan under the name of Taheebo NFD Marugoto (TNM), is expected to provide superior results to their original Taheebo due to its reduced particle size and greater aqueous solubility.

In an effort to evaluate the growth inhibitory effects and anti-aromatase activity of TNM, the experiments in the present study were designed to i) examine the growth inhibitory efficacy of TNM in a model for aromatase-expressing post-menopausal breast cancer, ii) evaluate the effects of TNM on cellular aromatase activity, and iii) identify possible molecular mechanisms responsible for the efficacy of TNM.

Materials and methods

Experimental model. The MCF-7AROM cell line represented the experimental model for the present study. These ER+/PR+/HER-2 human mammary carcinoma-derived cells, stably transfected with the aromatase gene (6,10), possess the characteristics of aromatase-expressing, post-menopausal Luminal A molecular subtype of clinical breast cancer.

Test compounds

Taheebo NFD Marugoto (TNM). This compound is comprised of finely ground powder from the inner bark of TA tree containing the bioactive agent Naphthofurandione (NFD) was provided by Taheebo Japan Co., Ltd.. The non-fractionated aqueous stock solution was prepared following the protocol provided by the supplier. This stock solution of TNM contains 200 ng of NFD (Personal Communication: Dr Fukuda, Taheebo Japan). The stock solution was serially diluted in the culture medium to obtain final concentrations of TNM containing 0.33% agar, and this cell suspension was overlaid on the basement layer in the presence or absence of TNM. The cultures were incubated at 37°C in a CO2 incubator for 21 days. The anchorage-independent (AI) colonies were stained with 0.005% crystal violet and colony counts were determined at x10 magnification. The data were expressed as number of AI colonies.

Cell cycle progression. For the cell cycle analysis, 5x10^5 cells were seeded in T-25 flasks and treated for 24 h. post-seeding with 1, 5 and 10 µg of TNM for 48 h. The cells were harvested by trypsination, pelleted at 500 x g, and washed twice with cold PBS (Sigma-Aldrich; Merck KGaA). An aqueous extract of Taheebo NFD Marugoto (TNM), is expected to provide superior results to their original Taheebo due to its reduced particle size and greater aqueous solubility.

Letrozole (LET). Stock solution of LET (molecular mass: 285 kDa, Sigma-Aldrich; Merck KGaA) was prepared in DMSO and serially diluted in the culture medium to obtain the final concentration of 1 µM (285 ng).

Exemestane (EXM). Stock solution of EXM (molecular mass: 296 kDa, Sigma-Aldrich; Merck KGaA) was prepared in DMSO and was serially diluted in the culture medium to obtain the final concentration of 10 µM (2,960 ng).

LET and EXM represent the prototypical aromatase inhibitors. The concentrations of 1 µM LET and 10 µM EXM are comparable to the effective concentrations traditionally used in the cell culture experiments, and represent clinically achievable effective concentrations. These compounds were used as positive controls for the present experiments.

Anchorage-independent growth. For this assay the stock solution of agar was prepared by mixing DNA grade agar (Sigma-Aldrich; Merck KGaA) with an appropriate volume of 2X RPMI-1640 medium. To prepare the basement layer, this stock solution was diluted to 0.6%, dispersed in a 6-well plate and allowed to solidify overnight at 37°C. Suspension of MCF-7AROM cells, at a density of 5x10^5 per ml, was prepared in RPMI-1640 medium containing 0.33% agar, and this cell suspension was overlaid on the basement layer in the presence or absence of TNM. The cultures were incubated at 37°C in a CO2 incubator for 21 days. The anchorage-independent (AI) colonies were stained with 0.005% crystal violet and colony counts were determined at x10 magnification. The data were expressed as number of AI colonies.

Caspase activity. Caspase 3/7 activity in the MCF-7AROM cells was measured using caspase-Glo assay kit (Promega). Briefly, the cells treated with TNM were homogenized by sonication in homogenization buffer (25 mmol/l HEPES, pH 7.5, 5 mmol/l MgCl2, and 1 mmol/l EGTA) and protease inhibitors (all from Sigma-Aldrich; Merck KGaA). The homogenate was centrifuged at 6,500 x g at 4°C for 15 min, and the supernatant was collected. Subsequently, 10 µl of assay reagent was added to 10 µl of supernatant and the reaction mixture was incubated at room temperature for 2 h. Resulting luminescence was measured using a Fluoroskan Luminometer (Thermo Scientific Co.). The data were expressed as relative luminescent units (RLU).

Gene expression profiling. The effect of TNM on the expressions of apoptosis regulatory genes BAX and BCL-2, E2 regulatory target genes ESR-1, AROM and PR and E2 responsive target genes pS2, GRB-2 and cyclin D1 was examined using reverse transcription quantitative PCR (RT-qPCR).
Table I. Primer sets used for reverse transcription-quantitative PCR analysis.

<table>
<thead>
<tr>
<th>Gene name</th>
<th>Primer sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ESR-1</strong></td>
<td>5'-TGTGCAATGACTGCTCCTCA-3' (S) 5'-GCTCTTCTCCTGTTTATA-3' (AS)</td>
</tr>
<tr>
<td><strong>AROM</strong></td>
<td>5'-AGCATGTCTAGCCAGCTGTT-3' (S) 5'-TCTCATACATTGGCCATGT-3' (AS)</td>
</tr>
<tr>
<td><strong>PR</strong></td>
<td>5'-ACGAAATTCTAGGCCGCTGTCGGG TGCAA-3' (S) 5'-ACAAGATCTCCACCGAGCCCG AGGTTT-3' (AS)</td>
</tr>
<tr>
<td>pS2</td>
<td>5'-CATCGACGGTCCTCCAGAAGAG-3' (S) 5'-CTCTGGGACTAATCACCCTGTG-3' (AS)</td>
</tr>
<tr>
<td><strong>GRB2</strong></td>
<td>5'-AAATGCTGCAAACACAGCGG-3' (S) 5'-TGAAGTCTGCGACATATCCTCC-3' (AS)</td>
</tr>
<tr>
<td><strong>Cyclin D1</strong></td>
<td>5'-ACGAAGGCTTGCGGTGT-3' (S) 5'-CCGCTGGCCTGAACTACCT-3' (AS)</td>
</tr>
<tr>
<td><strong>BCL-2</strong></td>
<td>5'-CTCTGGGACTAATCACCCTGTG-3' (S) 5'-TGAAGTCTGCGACATATCCTCC-3' (AS)</td>
</tr>
<tr>
<td>BAX</td>
<td>5'-GTTCATCCAGATGCAGGACGAG-3' (S) 5'-ATCCCTTTCTCTCAGATGTTGA-3' (AS)</td>
</tr>
<tr>
<td>β-actin</td>
<td>5'-GACCCTCTATGCAACACAGT-3' (S) 5'-AGTACTTGGCTCAGGGAGA-3' (AS)</td>
</tr>
</tbody>
</table>

Aromatase activity. To measure aromatase enzymatic activity, the $^3$H$_2$O release assay was used. [1H] androstenedione represented the substrate and its conversion to E$_2$ represented a measure for aromatase activity. The MCF-7AROM cells grown in phenol-free RPMI-1640 medium supplemented with charcoal-stripped fetal bovine serum (Sigma-Aldrich; Merek KGaA) were suspended in an assay mixture containing 0.1% bovine serum albumin, 67 mmol/l KPO$_4$ (pH 7.4), and 2.0 µmol/l progesterone. After sonication, 100 nmol/l of [1H] androstenedione (25.3 ci/mmol, NET-962; Perkin-Elmer, Inc.) was added and the mixture was incubated for 10 min at room temperature. NADPH was then added to a final concentration of 1.2 mmol/l, followed by 37°C incubation and the addition of an equal volume of 5% trichloroacetic acid. The supernatant was collected and extracted with an equal volume of chloroform. Dextran-coated charcoal was added to the assay mixture, which was then vortexed and centrifuged at 6,500 x g at 40°C for 15 min. The supernatant was then added to scintillation fluid and measured in a scintillation counter (Perkin-Elmer). The data were expressed as f mole E$_2$ formed, per mg protein, per hour (21).

Statistical analysis. The experiments for dose response using the anchorage-independent growth assay were conducted in quadruplicate. Experiments for cell cycle progression, caspase 3/7 activity, aromatase activity and gene expression profiling were conducted in triplicate. The data were expressed as mean ± SD. Statistically significant differences between the control and treatment groups were assessed by the two-sample Student's t-test. P<0.05 was considered to indicate a statistically significant difference. Additionally, data from comparisons of multiple treatment groups were analyzed using analysis of variance (ANOVA) and Dunnett's test as a post-hoc test with a threshold of α=0.05 (Microsoft Excel 2013 XLSTAT-Base software).

Results

Effect of TNM on anchorage-independent growth. The data presented in Fig. 1 show a TNM dose-dependent reduction in the number of E$_2$ promoted anchorage-independent colonies.
These data identified the IC₅₀ as 5 µg TNM (α=0.05), and IC₉₀ as 20 µg TNM (α=0.05), relative to the E₂-treated controls. The NFD content of TNM at these concentrations was estimated to be 1.0 and 4.0 ng, respectively.

**Effect of TNM on cell cycle progression.** The data presented in Table II examined the effect of TNM on the cell cycle progression of MCF-7AROM cells. TNM at the maximum cytostatic concentration of 10 µg resulted in 62.2% of cells arrested in the S phase of the cell cycle (P=0.04), relative to the untreated control. The inhibition in the G₁ and G₂ phases were modest and statistically non-significant.

**Induction of pro-apoptotic activity by TNM.** The induction of cellular apoptosis by treatment with TNM was examined by monitoring the status of caspase 3/7 activity. In response to treatment with TNM, caspase 3/7 activity exhibited a dose-dependent increase (Fig. 2A). Thus, relative to the untreated control, TNM at 1.5 µg, 5 µg and 10 µg exhibited a 2.9-fold (α=0.05), an 8.9-fold (α=0.05) and a 9-fold (α=0.05) increase in caspase 3/7 activity, respectively.

At the molecular level, TNM treatment resulted in a dose-dependent increase in pro-apoptotic BAX and a reciprocal decrease in anti-apoptotic BCL-2 gene expression. Thus, treatment with 10 µg TNM resulted in a 3.3-fold increase (P=0.01) vs. untreated control. BAX: *P=0.01 vs. untreated control. BCL-2: *P=0.02 vs. untreated control. TNM; Taheebo NFD Marugoto, BAX; BCL-2 associated X protein; BCL-2; B cell lymphoma-2; SD, standard deviation.

**Inhibition of aromatase activity by TNM.** In response to treatment with TNM, MCF-7AROM cells exhibited dose-dependent inhibition in aromatase activity as measured by the extent of conversion of androstenedione to E₁. Thus, TNM treatment at 30, 40 and 50 µg resulted in a 42% (α=0.05), a 57.9% (α=0.05) and a 97.1% (α=0.05) decrease in the aromatase activity, relative to the untreated control (Fig. 3).
Comparative inhibition of aromatase activity by TNM, LET and EXM. The comparative efficacy of TNM and LET for inhibition of aromatase activity revealed that the extent of inhibition of 63.8% (P=0.04) by 40 µg TNM (NFD content 8 ng) was essentially similar to 62.3% inhibition (P=0.04) induced by 285 ng (1 µM) of LET, relative to the untreated control (Table III).

The comparative efficacy of TNM and EXM for inhibition of aromatase activity is presented in Table IV. These data revealed that the extent of inhibition of 98.5% (P=0.01) by 100 µg TNM (NFD content 20 ng) was essentially similar to 97.1% inhibition (P=0.01) induced by 2,960 ng (10 µM) of EXM, relative to the untreated control.

Inhibition of E₁ regulated target gene expression by TNM. The data obtained from the effect of TNM on the expression of ESR-1 (gene for ER-α), AROM and PR genes are presented in Fig. 4A. The extent of inhibition at 10 µg of TNM for ESR-1 was 90% (P=0.01), for AROM it was 61% (P=0.04) and for PR it was 61% (P=0.04). Thus, treatment with TNM resulted in substantial downregulated expressions of select genes that are regulated by E₁.

Inhibition of E₂ responsive target gene expression by TNM. The data shown in Fig. 4B examined the effect of TNM on the expression of select E₂ responsive genes. The extent of inhibition at 10 µg of TNM for pS2 was 62% (P=0.04), for GRB2 it was 61% (P=0.04), and for cyclin D1 it was 82% (P=0.01). Thus, TNM treatment resulted in a substantial downregulation of E₂ responsive gene expressions.

Discussion

Metastatic breast cancer is a leading cause of cancer related mortality for women in the USA (22). The ER-α positive, aromatase-expressing Luminal A subtype of post-menopausal breast cancer responds to aromatase inhibitors (6,10). However, long-term therapy is frequently associated with acquired resistance that negatively impacts efficacy and facilitates disease progression.

In addition to the present MCF-7AROM model, other cellular models stably transfected with the aromatase gene have been developed from human mammary carcinoma derived MCF-7 and T47D cell lines. These models have been utilized to examine the effects of aromatase inhibitors and investigate the mechanisms responsible for resistance to AI-based endocrine therapy. For example, MCF-7AROM cells have exhibited resistance to Fluvestrant and cross resistance to Letrozole, Anastrazole, Exemestane (9) and T47DAROM cells have exhibited resistance to Letrozole and sensitivity to anti-progestin (23).

Collectively, these three cellular models offer valuable experimental approaches to identify efficacious aromatase inhibitors and also to investigate molecular mechanisms responsible for acquired resistance to aromatase inhibitors.

Non-toxic natural nutritional products may represent testable alternatives for endocrine therapy-resistant post-menopausal breast cancer (17,18, 28-31), and thereby, may provide treatment options against the clinical limitations of current aromatase inhibitor-based therapy (2,3,6,9,10). Two species belonging to the Tabebuia genus have documented anti-cancer activity. The anti-cancer effects of T. avellanedae and T. chrisantha are documented in preclinical xeno-transplant models (13) and in mice carrying Ehrlich ascites tumor (24). The use of TA in traditional medicine is not well documented. However, the effect of an aqueous extract of TA has been examined on the status of quality of life in patients with multiple organ site cancers that are at advanced metastatic stages (25). Additionally, the effects of the TA quinone NFD have been

Table III. Comparative efficacy for aromatase inhibition by TNM and LET in MCF-7AROM cells.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Concentration</th>
<th>Aromatase activity (E₁ fmole/mg protein/h)</th>
<th>Inhibition (% control)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>-</td>
<td>6.9±0.4a</td>
<td>-</td>
</tr>
<tr>
<td>TNM (40 µg)</td>
<td>8 ng NFD</td>
<td>2.5±0.1b</td>
<td>63.8</td>
</tr>
<tr>
<td>LET (1 µM)</td>
<td>285 ng</td>
<td>2.6±0.1b</td>
<td>62.3</td>
</tr>
</tbody>
</table>

Results were expressed as mean ± SD, n=3 per treatment group. *P<0.05. Data were analyzed by the two-sample Student’s t-test. E₁, estrone; TNM, Taheebo NFD Marugoto; LET, Letrozole; NFD, naphthofurandione; SD, standard deviation.

Table IV. Comparative efficacy for aromatase inhibition by TNM and EXM in MCF-7AROM cells.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Concentration</th>
<th>Aromatase Activity (E₁ fmole/mg protein/h)</th>
<th>Inhibition (% control)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>-</td>
<td>6.9±0.30a</td>
<td>-</td>
</tr>
<tr>
<td>TNM (100 µg)</td>
<td>20 ng NFD</td>
<td>0.1±0.08b</td>
<td>98.5</td>
</tr>
<tr>
<td>EXM (10 µM)</td>
<td>2,960 ng</td>
<td>0.2±0.10b</td>
<td>97.1</td>
</tr>
</tbody>
</table>

Results were expressed as mean ±SD, n=3 per treatment group. *P<0.05. Data were analyzed by two-sample Student’s t-test. E₁, estrone; TNM, Taheebo NFD Marugoto; EXM, exemestane; NFD, naphthafurandione; SD, standard deviation.
At the mechanistic levels, the anti-proliferative effects of TNM were evidenced by induction of S-phase arrest and resultant inhibition of cell cycle progression. The pro-apoptotic effects of TNM were evidenced by the dose-dependent induction of caspase 3/7 activity. The pro-apoptotic BAX and anti-apoptotic BCL-2 genes are critical for the mitochondrial intrinsic apoptosis (32). TNM treatment resulted in a dose-dependent increase in the expression of pro-apoptotic BAX gene and decrease in the expression of anti-apoptotic BCL-2 gene. These data exhibiting reciprocal modulation of mRNA expression for these genes provide mechanistic leads that support the pro-apoptotic effect of TNM in the present MCF-7AROM model. In the E2-mediated signal transduction pathways pS2, GRB2 and cyclin D1 represent classical E2 responsive target genes (11,12). Thus, collectively, the inhibitory effects of TNM on E2 regulated ESR-1 (gene for ER-α), PR and AROM genes and on E2 responsive pS2, GRB2 and cyclin D1 genes provide mechanistic leads relevant to possible molecular targets for the efficacy of TNM via the ER signal transduction pathways.

Long-term treatment with the pharmacological inhibitors of aromatase is frequently associated with systemic toxicity and acquired drug resistance (2,3,6,10) leading to the emergence of therapy resistant stem cells. By contrast, naturally occurring TNM may exhibit lower systemic toxicity and may lack drug resistance that is induced by pharmacological inhibitors of aromatase activity. Inhibitory effect of TNM on aromatase activity is evidenced by its ability to reduce the conversion of androstenedione to E2 in MCF-7AROM cells in a dose-dependent manner. The specificity of this effect is indicated by the induction of aromatase inhibition by two selective pharmacological aromatase inhibitors LET and EXM. In this context, it is noteworthy that TNM at a concentration of 4 µg (NFD content: 8 ng) exhibits aromatase inhibition that is essentially comparable to 285 ng of LET. Therefore, based on the NFD content of TNM, it requires 35.6-fold higher concentration of LET which is indicative of a 35.6-fold greater potency of TNM. Additionally, TNM at a concentration of 100 µg (NFD content: 20 ng) exhibits aromatase inhibition, which is essentially comparable to 2,960 ng of EXM. Therefore, based on the NFD content of TNM, it requires 148-fold higher concentration of EXM which is indicative of a 148-fold greater potency of TNM. It is also noteworthy that extracts from natural products such as resveratrol and ellagitannin derivatives have documented anti-aromatase activity (33,34). In addition, natural products such as sulforaphane, benzyl isothiocyanate, a vitamin A derivative all-trans retinoic acid and a terpenoid carnosol have been documented to target cancer stem cells (35-37).

The present data identified several potential mechanistic leads responsible for TNM-mediated anti-aromatase activity. For example, the clinical aromatase inhibitor LET binds the active site of the aromatase enzyme, and EXM functions as a substrate analogue for the enzyme (1). TA downregulates ESR-1 and E2 metabolizing enzymes CYP1A1 and CYP1B1 (17), and ER-α, and as its ligand E2 induces aromatase expression (38). Thus, TNM-mediated inhibition of the ER-α gene ESR1 and of AROM raise the possibility that TNM may be effective via one or more of the mechanisms discussed above.

At present, direct evidence to support efficacy of the active principle in TNM is equivocal. However, published evidence has demonstrated anti-cancer activity of NFD in animal...
models (13). In addition to NFD, another quinone β-lapachone (β-LAP) has been documented to be present in trace amounts in the non-fractionated aqueous extracts of TA and TNM. This minor constituent, at higher pharmacological concentrations, exhibits anti-cancer activity in preclinical xenotransplant models of epithelial organ site cancers via distinct molecular mechanisms (15,16,39-41). However, it is important to recognize that at the low concentrations of TA or TNM used, the levels of β-LAP remain essentially undetectable (42).

The data from the present study have identified several potential mechanistic leads for the inhibitory efficacy of TNM in cellular models of human cancer. These leads include modulation of the RB signaling pathway, inhibition of cyclin-dependent kinase, inhibition of Cdc dual phosphatase, inhibition of cyclo-oxygenase-2, inhibition of telomerase and downregulated global expression of several genes that are involved in cell cycle progression, cellular apoptosis and hormone metabolism (14-18). Thus, collectively, these lines of evidence provide potential leads that the efficacy of TNM in the present model is likely due to its NFD content.

In conclusion, the data presented in this study, provide evidence for the growth inhibitory activity of TNM in a cellular model for aromatase-expressing post-menopausal breast cancer. More importantly, these data identify mechanistic leads as a proof of concept that TNM may be a superior naturally occurring substitute for clinical aromatase inhibitors. Overall, the present study validates an experimental approach for mechanistic evaluation of additional natural products that exhibit anti-aromatase activity. In this context, strong mechanistic lead for the efficacy of TNM in the present cellular study reveal that future experimental approaches that are designed to provide clinically translatable therapeutic evidence for the use of TNM. These approaches may include experiments on the MCF-7AROM tumour model to examine the effects of TNM on tumor progression and on molecular characteristics of tumors relevant to altered E2-mediated signal transduction pathways. Additionally, for clinical translatability of the present preclinical study, experiments to obtain clinical data on absorption, distribution, metabolism and excretion (ADME) of TNM, and on data for human safety, tolerability and efficacy of TNM may provide valuable information.

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Availability of data and materials

The data sets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors’ contributions

NT conceived the study design, formulated the experimental protocols, and prepared the manuscript. HBN conducted all the experiments, organized and analyzed the data and participated in the preparation of the manuscript. GYCW selected the test agent and contributed to data interpretation and preparation of the manuscript. All authors have read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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